# <u>LETTERS</u>

# Highly Rigid Labdane-Type Diterpenoids from a Chinese Liverwort and Light-Driven Structure Diversification

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**(5)** Supporting Information

**ABSTRACT:** Two unprecedented labdane-type diterpenoids haplomintrins A (1) and B (2) with six rings system were isolated from a Chinese liverwort *Haplomitrium mnioides*. Light-driven reaction of homologous haplomitrenonolides C (6), A (4), and D (3) afforded haplomintrins A–C (1, 2, and 7), respectively, while 4 was converted to more complex congeners haplomintrins D–G (8–11) through intramolecular cyclization. Formation of 1 and 2 from compounds 6 and 4, respectively, helps us to postulate that a photochemical reaction is involved in the biosynthetic pathway. These structure features can be used as molecular markers of *H. mnioides*, and their allelopathic effects are also preliminarily tested.

As the most primitive group of terrestrial plants, liverworts are well-known natural resources for structurally diverse terpenoids and aromatic compounds that exhibit interesting biological properties, such as cytotoxic, antifungal, allelopathic, insect antifeedant, and antioxidantive effects.<sup>1</sup> The liverwort Haplomitrium mnioides (Lindb.) R. M. Schust is considered as a very primitive taxon,<sup>2</sup> with remote affinity to other liverwort groups.<sup>3</sup> Previous chemical investigation on this species produced several chain-type and labdane-type diterpenoids.<sup>3b</sup> Our continued study on the labdane-typediterpenoids from Chinese liverworts<sup>4</sup> led us to discover two novel compounds, haplomitrins A and B (1 and 2), possessing unprecedented scaffolds, one new biosynthetically related labdane haplomitrenolide D (3), as well as three known haplomitrenolides A–C  $(4-6)^{3b}$  from *H. mnioides* (Figure 1). Light-driven reaction of compound 6 to 1 and 4 to 2, respectively, enabled us to speculate that the photochemical reactions are involved in the late step of the biosynthetic pathway of these diterpenoids. The formation of more complex congeners haplomintrins C-G (7– 11) during the photochemical reaction provides one useful way to diversify the natural products.

Haplomitrin A (1) was obtained as a colorless needle. Its molecular formula of  $C_{21}H_{24}O_6$  was established by HRESIMS  $(m/z \ 373.1653 \ [M + H]^+$ , calcd  $C_{21}H_{25}O_6$ , 373.1646), requiring 10 indices of hydrogen deficiency. Analysis of the <sup>1</sup>H NMR data (Table 1) revealed the existence of two tertiary methyls at  $\delta_H$  1.72 (s) and 1.24 (s), an oxygenated methyl group at  $\delta_H$  3.69 (s), and one disubstituted double-bond group ( $\delta_H$  6.52, d, J = 2.8 and 4.77 d, J = 2.8). The <sup>13</sup>C NMR (Table 1) and HMQC data showed 21 carbons, including one ketone carbonyl ( $\delta_C$  204.8), two ester carbonyls ( $\delta_C$  172.3 and 179.4), four quaternary carbons, six methines (two oxygenated, two





**Figure 1.** Structures of labdane-type diterpenoids 1-6 isolated from *H. mnioides* and congeners 7-11 formed by photochemistry reaction.

olefinic and two other), four methylenes, three methyls (one oxygenated). The aforementioned data suggested a modified labdane framework with a  $\delta$ -lactone ring and a methyl ester carbonyl. Compared with the known structure of haplomitrenonolide C (6) isolated from the same plant,<sup>3b</sup> the disappearance of the two double bonds  $\Delta^{7(8)}$  and  $\Delta^{13(16)}$  in the <sup>13</sup>C NMR of 1 and the appearance of the coupling correlations between H-7 ( $\delta_{\rm H}$  2.91) and H-16 ( $\delta_{\rm H}$  4.72) in <sup>1</sup>H–<sup>1</sup>H COSY demonstrated the formation of a new tetracycle.

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	1			2
no.	$\delta_{ m C}$	$\delta_{ m H}$	$\delta_{ m C}$	$\delta_{ m H}$
$1\alpha$	38.6 t	1.39 m	39.5 t	1.38 td (13.6, 3.6)
$1\beta$		2.39 br d (13.6)		2.40 br d (13.6)
$2\alpha$	17.6 t	1.62 m	18.4 t	1.49 m
2β		2.10 ddd (14.0, 3.2, 3.2)		2.00 m
3α	37.9 t	1.45 m	42.9 t	1.13 m
3β		1.82 m		1.50 m
4	43.5 s		32.7 s	
5	59.4 d	3.03 s	63.7 d	1.94 s
6	204.8 s		206.6 s	
7	56.4 d	2.91 d (3.2)	57.5 d	2.83 d (3.2)
8	50.1 s		50.7 s	
9	55.3 d	2.64 dd (4.8, 2.0)	56.0 d	2.04 dd (4.4, 1.2)
10	47.1 s		48.7 s	
11α	33.7 t	2.33 d (13.6)	33.9 t	2.30 d (13.2)
$11\beta$		1.86 m		1.79 ddd (13.2, 4.8, 2.8)
12	83.1 d	4.55 br s	83.2 d	4.54 br s
13	66.8 s		66.6 s	
14	100.2 d	4.77 d (2.8)	100.1 d	4.71 m
15	150.7 d	6.52 d (2.8)	150.9 d	6.52 d (2.8)
16	80.3 d	4.72 d (3.2)	81.7 d	4.72 m
17	23.0 q	1.24 s 3H	23.5 q	1.23 s 3H
18	179.4 s		34.2 q	1.08 s 3H
19	16.5 q	1.72 s 3H	21.1 q	1.47 s 3H
20	172.3 s		172.5 s	
COOMe	52.6 g	3.69 s 3H		

Table 1.	<sup>1</sup> H (	(400 MHz)	) and <sup>13</sup> C	(100 MHz	) NMR Data	(in ppm	) for 1	1 and 2 in CDCl <sub>3</sub>
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HMBC correlations from H-9 ( $\delta_{\rm H}$  2.64) to C-13 ( $\delta_{\rm C}$  66.8) and from H-12 ( $\delta_{\rm H}$  4.55) to C-8 ( $\delta_{\rm C}$  50.1) also confirmed the linkages (Figure 2A). Thus, compound **1** was elucidated as a highly rigid structure with a novel tetracyclo[7.4.1.0<sup>2,7</sup>.0<sup>11,14</sup>]tetradecane skeleton.



Figure 2. (A) Selected HMBC  $(H \rightarrow C)$  and  ${}^{1}H^{-1}H \text{ COSY }(H-H)$  correlations of 1. (B) Selected NOESY correlations  $(H \leftrightarrow H)$  of 1.

NOESY correlations (Figure 2B) of H-5/H-9, H-9/H<sub>3</sub>-17, H<sub>3</sub>-17/H-14, and H-7/H<sub>3</sub>-17 indicated the relative configuration of **1**. To determine the absolute configuration of **1**, a single-crystal X-ray diffraction analysis with Cu K $\alpha$  radiation was performed (CCDC 1059512, Figure 3). Accordingly, the stereochemistry of **1** was determined as 4*R*,5*S*,7*S*,8*R*,9*S*,10*S*,12*R*,13*S*,16*R*.

The same strategy was adopted for the elucidation of the structure of **2**, based on its molecular formula of  $C_{20}H_{24}O_4$ , as determined by HRESIMS (m/z 329.1752 [M + H]<sup>+</sup>, calcd 329.1747) and comparison of its NMR data with those of **1** (Table 1), which showed the absence of resonances for the methyl ester carbonyl and the appearance of an additional tertiary methyl at C-18 ( $\delta_{\rm H}$  1.08,  $\delta_{\rm C}$  34.2) in **2**. This deduction was confirmed by the HMBC correlations from H<sub>3</sub>-18/H<sub>3</sub>-19



Figure 3. X-ray crystallographic structures of 1 and 9.

to C-3, C-4, and C-5. Therefore, the molecular structure of 2 was established as shown.

Compound 3 was also a new compound which had a molecular formula of  $C_{21}H_{24}O_7$ , with 10 degrees of unsaturation, as established by HRESIMS data. Analysis of its 1D and 2D NMR data (Supporting Information) determined 3 to be the 9-hydroxyl derivative of haplomitrenonolide C (6). Compounds 4 and 5 were known and had never isolated from the same plant.<sup>3b</sup>

Topologically, the novel labdane-type diterpenoids show very special features. They have a six-ring system with an untouched six-membered latctone ring as well as a fused furan ring. Obviously, these unique structures are formed through intramolecular cyclization. Here, we outlined a plausible biosynthetic pathway of these novel compounds (Scheme 1A). Intramolecular [2 + 2] cycloaddition<sup>5</sup> of the precursor **6** enzymatically or nonenzymatically delivered the highly rigid structure of **1**, while compound **4** converted to **2** by a same mechanism. Also, an alternative way to form **1** from **2** was through C-18 methyl oxidation into a carboxylic acid followed by a methyl esterification.

Scheme 1. (A) Proposed Biosynthetic Pathways of Compounds 1 and 2. (B) Proposed Mechanism of Photodriven Formation of Compounds 8-11



According to the above analysis, it is reasonable to speculate that a photochemical-catalyzed intramolecular cyclization was involved in the formation of 1 and 2 biosynthetically<sup>o</sup> as they are not artifacts as detected in the crude extract of the fresh material (Figure S33). As expected, irradiation of 6 with UV light (low pressure Hg lamps, 400 W, 254 nm) under nitrogen atmosphere at room temperature afforded compound 1(31% yield), as detected by the time-lapse HPLC monitoring analysis (Figure S35). Applying the same protocol to 4 and 3 generated 2 (34% yield) and 7 (33% yield), respectively (Figure S36 and S37). The <sup>1</sup>H and <sup>13</sup>C NMR data (Table S2) of 7 suggested the disappearance of two double bonds. 2D NMR data (Supporting Information) confirmed that an intramolecular  $\begin{bmatrix} 2 \\ + 2 \end{bmatrix}$ photocycloaddition has arisen between the  $\Delta^{7(8)}$  and  $\Delta^{13(16)}$ double bonds with good diastereoselectivity. Compound 7 was determined to be the 9-OH derivative of 1, which was not found in the crude plant extract by HPLC analysis (Figure S33). In addition, the increase in the contents of 1 and 2 when the plant species was exposed to prolonged daylight also supports the above speculation (Figure S34).

Surprisingly, these complex labdane-type diterpenoids were not the end products of the photochemical reaction cascade. In the light-driven reaction, several other photoadducts were detected by the HPLC traces. In order to establish the structures, the amplified reaction of **4** was subjected to the irradiation, and after all of the starting material was consumed (Figure S51), a mixture of compound **2** and compounds **8–11** (29% yield, **2/8/9/10/11** = 3:2:4:3:2) was obtained.

To determine the structures of these photochemical products, NMR spectroscopy and X-ray diffraction analysis were employed. Compound 8 was assigned the molecular formula of  $C_{20}H_{24}O_4$  by HRESIMS data (Figure S60). The 2D NMR data (Table S3) of 8 were close to those of 4, except for

the absence of C-18 methyl and the C-6 ketone carbonyl in 4 and the presence of a methylene group ( $\delta_{\rm H}$  2.20, m, 2H;  $\delta_{\rm C}$  55.90) in 8. Its molecular structure with a tricyclo[3.5.1.0<sup>3,11</sup>]-undecane skeleton was confirmed on the basis of the HMBC data (Figure S52). The methyl at C-4 was determined to be  $\beta$  oriented, supported by the NOESY correlations (Figure S52) of H<sub>3</sub>-19/H-1 $\beta$  and H-18/H-5. The hydroxyl group at C-6 revealed a  $\beta$  configuration from the stereoconsideration in which *cis* arrangement of the tetratomic ring was required.

Compound 9 was obtained as a colorless crystal with a molecular formula  $C_{20}H_{24}O_4$ , as established by HRESIMS (Figure S72). Compound 9 was confirmed to be the cagelike labdane possessing an unprecedented pentacyclic [7.4.1.1<sup>2,4</sup>.0<sup>8,15</sup>.0<sup>12,14</sup>] pentadecane scaffolding bearing two heterocylic rings by comparing the 1D and 2D NMR data (Table S4 and Figure S64) with those of 2. The relative configuration of 9 was verified by the NOESY correlations (Figure S65). Single-crystal X-ray diffraction study permitted the unambiguous assignment of its absolute configuration (CCDC 1059598, Figure 3).

Compounds 10 and 11 were determined to be a pair of diastereoisomers. Their identical molecular formula  $(C_{20}H_{24}O_4)$  was established by HRERMS (Figures S84 and S94). Aside from the absence of resonances for the  $\Delta^{14(15)}$  double bond and the presence of aldehyde groups at C-15 ( $\delta_{\rm H}$  9.90,  $\delta_{\rm C}$  198.2 in 10 and  $\delta_{\rm H}$  9.04,  $\delta_{\rm C}$  196.5 in 11), other assignments of NMR data (Table S5) resembled with those of 2. Moreover, the newly formed cyclopropane ring was confirmed by the connectivity of -C-7(H)-C-16(H)-C-14(H)- established by the <sup>1</sup>H-<sup>1</sup>H COSY spectrum and the HMBC correlation from H-14 to C-15 (Figure S76), elucidating the striking C-14/C-16 linkage. Therefore, the unique pentacyclic [7.4.1.0<sup>2,7</sup>.0<sup>10,12</sup>.0<sup>12,15</sup>]pentadecane frameworks of 10 and 11 were determined. According to their NOESY data (Figure S77), the aldehyde groups of 10 and 11 were determined respectively.

To investigate the formation of these unique carbon skeletons formed by a photochemical reaction, we proposed that the complexity was achieved through photoexcited diradical intermediates (Scheme 1B). Compound 8 arose from 4 and 9 from 2, in which formation of the cyclobutane ring stereoselectively was through the Norrish–Yang cyclization<sup>5a,7</sup> with 1,4-diradical 13 and 14 as intermediates, respectively. In addition, the cleavage of the furan ring of 2 at C16–O forming 10 and 11 was postulated through a diradical intermediate 15. This is the first report related to the light-induced furan ring opening to form a cyclopropyl ring attached with an aldehyde group.

Ecologically, besides the phenological responses, the chemicals metabolized by liverworts also help them to cope with their living conditions by combatting other organisms.<sup>8</sup> Here, we tested the allelopathic activity of the isolated compounds **2** and **4**, the representives of two skeletons, with the model of *Arabidopsis thaliana*. The results showed that compounds **2** and **4** could regulate the growth of *A. thaliana* by inhibiting the root elongation with the IC<sub>50</sub> at 44.57 ± 0.78 and 19.08 ± 0.73 µg/mL respectively (Figure S107).

In conclusion, haplomitrins A and B (1 and 2) are rearranged labdane-type diterpenoids with unprecedented scaffolds isolated from Chinese liverwort. Their structures had been clarified, and the formation of 1 and 2 from 6 and 4 through light-driven [2 + 2] cycloaddition was demonstrated. Formation of haplomintrin C (7) from 3 and haplomintrins D-G (8–11) with more appealing labdane skeletons from 4 after an extended amount of time through light-driven conversion provides one intriguing way to diversify natural products. Finally, phytotoxic activities of typical compounds have been preliminarily studied. Taken together, our work shows the UV/vis light involving in the biosynthesis step in vivo or photodriven transformation in vitro will access to more complex structures.

## ASSOCIATED CONTENT

#### **S** Supporting Information

General experimental procedures, structure elucidation of **3** and 7–11, 1D and 2D NMR data, HRESIMS, IR, UV, and CD spectra of the new compounds, 1D NMR data and ESIMS of the known compounds, crystal information for **1** and **9**, HPLC and UV analysis for the photoinduced reaction, and ecological biochemistry tests. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/ acs.orglett.5b01664.

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#### notes

The authors declare no competing financial interest.

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